



QUIDEL®

Technical Bulletin

Decontamination and Contamination Prevention

Avoiding contamination is one of the most important steps for any RNA or DNA amplification testing. Prior to processing specimens, disinfect surfaces with a 1% bleach solution¹ (which contains 0.05% chlorine concentration). More diluted bleach solutions are not effective. The chlorine must subsequently be removed as it is caustic and may damage equipment. This may be done either by wiping the surfaces with clean water or with 70% alcohol (which also has a useful, additional effect against most bacteria [not against bacterial spores] and vegetative fungi). **A 10% bleach solution, which contains a 0.5% chlorine concentration, is recommended for decontamination of blood or body fluid spills.**¹

Specimens should be handled one at a time without having multiple tubes opened at once. Specimens should be carefully transferred from primary containers using disposable pipettes. If it is suspected that micropipettes have been contaminated during the transfer or testing process, immediately discontinue use of the pipette. Contaminated pipettes should be thoroughly cleaned using the same process as described above for the work surfaces. If gloves are thought to be contaminated at any point during testing, they should be changed immediately before proceeding to the next sample or continuing with the remainder of the procedure.

AmpliVue®

- Clean all surfaces such as bench tops, all work areas, heat blocks, and the specimen rack involved with AmpliVue testing with a lint-free tissue or cloth dipped in a 1% bleach solution, followed by wiping with water. **Repeat the procedure 3 times with 10% bleach for decontamination.** After the final wipe with water, 70% alcohol (isopropanol or ethanol) can be used to wipe off the remaining traces of bleach.
- Handle specimens one at a time without having multiple tubes opened at once.
- If it is suspected that the pipette utilized for testing has become contaminated, discontinue use immediately. Most pipettes can be cleaned with a bleach solution as described above; however, if there is any question about appropriate cleaning of the pipette, contact the pipette manufacturer. Extended length pipette tips can provide contamination prevention by allowing access to the sample without contact by the pipette.

¹ <http://www.who.int/csr/resources/publications/surveillance/Annex7.pdf>

To prepare 1:10 bleach solution add one volume of household bleach (e.g. 1 liter) to nine volumes of clean water (e.g. 9 liters).

To prepare 1:100 bleach solution add one volume of 1:10 bleach solution (e.g. 1 liter) to nine volumes of clean water (e.g. 9 liters).

Note: 1:100 bleach solution can also be prepared directly from household bleach by adding 1 volume of household bleach to 99 volumes of clean water (e.g. 100 ml of bleach to 9.9 liters of clean water) but making it up from 1:10 bleach solution is much easier).

- When gloves are suspected of being contaminated or are visibly soiled (i.e., liquid appears on the glove), discard and change immediately before further testing is performed.

Note: *Reaction Tubes or Detection Chambers should never be reopened after amplification to avoid potential amplicon contamination in the laboratory.*

Solana®

- Turn off and unplug the equipment/instrument before cleaning.
- Clean all surfaces such as bench tops, all work areas, heat block, workflow tray, and specimen transfer rack where Solana testing is performed with a lint-free tissue or cloth dipped in a 1% bleach solution, followed by wiping with water. **Repeat the procedure 3 times with 10% bleach for decontamination.** After the final wipe with water, 70% alcohol (isopropanol or ethanol) can be used to wipe off the remaining traces of bleach.
- Handle specimens one at a time without having multiple tubes opened at once.
- If it is suspected that the pipette utilized for testing has become contaminated, discontinue use immediately. Most pipettes can be cleaned with a bleach solution as described above; however, if there is any question about appropriate cleaning of the pipette, contact the pipette manufacturer. Extended length pipette tips can provide contamination prevention by allowing access to the sample without contact by the pipette.
- When gloves are suspected of being contaminated or are visibly soiled (i.e., liquid appears on the glove), discard and change immediately before further testing is performed.
- Pay special attention when cleaning the display and the tube holes. **Take care that no liquid runs onto the edges of the display, the tube holes or the connectors.**
- The entire surface should be wiped with a thin, moist film of disinfectant and then allowed to dry.

Note: *Reaction Tubes should never be reopened after amplification to avoid potential amplicon contamination in the laboratory.*

Note: *Instrument should not be cleaned with a spray bottle.*

Note: *Do not place any cleaning solutions into the tube wells.*

Note: *Do not pour cleaning solutions directly onto the instrument.*

Lyra®

- Clean all surfaces such as bench tops, all work areas, heat blocks, and any sample racks utilized for Lyra testing with a lint-free tissue or cloth dipped in a 1% bleach solution, followed by wiping with water. **Repeat the procedure 3 times with 10% bleach for decontamination.** After the final wipe with water, 70% alcohol (isopropanol or ethanol) can be used to wipe off the remaining traces of bleach.
- Handle specimens one at a time without having multiple tubes opened at once.
- If it is suspected that the pipette utilized for testing has become contaminated, discontinue use immediately. Most pipettes can be cleaned with a bleach solution as described above; however, if there is any question about appropriate cleaning of the pipette, contact the pipette manufacturer. Extended length pipette tips can provide contamination prevention by allowing access to the sample without contact by the pipette.
- When gloves are suspected of being contaminated or are visibly soiled (i.e., liquid appears on the glove), discard and change immediately before further testing is performed.

Note: *Reaction Tubes or PCR plates should never be reopened after amplification to avoid potential amplicon contamination in the laboratory.*

If contamination has occurred or false positive results are obtained with known negative patient samples or with any negative control, including the negative control that is included in any of the Quidel Molecular control sets, immediately decontaminate bench tops, heat blocks, pipettes, and any additional equipment or surfaces that may have been affected using a 10% bleach solution as described above. If false positive results still occur, contact our Technical Support department.

If you have any questions regarding the use of this product, please contact Quidel Technical Support at 1.800.874.1517 (in the U.S.) or technicalsupport@quidel.com. If outside the U.S., further information can be obtained from your distributor, or directly from Quidel at one of the numbers listed below. Reference quidel.com to see more options for Support.

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You may also visit our website at quidel.com for information on Quidel's line of Rapid Diagnostics, Molecular Diagnostics, Cell Culture and Specialty Products (Bone Health and Autoimmune & Complement). Other product information available on our website includes: CPT codes, CLSI procedure guides, SDS, and Package Inserts.

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