



MicroVue™ Bone

Mouse Osteocalcin EIA

For the determination of mouse osteocalcin levels in serum, plasma or cell culture media

For **Research Use Only** in the United States. Not for use in diagnostic procedures.

A symbols glossary can be found at quidel.com/glossary.

INTRODUCTION

Mouse osteocalcin, a 46 amino acid peptide, is the major noncollagen protein found in mouse bone. It contains three gamma- carboxyglutamic acid (GLA) residues at positions 13, 17, and 20 and is, therefore, also known as bone gla-protein or BGP. The exact biological function of osteocalcin is not known but the three gamma-carboxyglutamic acid residues confer on it a very strong ability to bind to hydroxyapatite and calcium.

Vitamin K is essential for the biosynthesis of osteocalcin which is stimulated by 1,25 -dihydroxyvitamin D. Osteocalcin is synthesized by osteoblasts during the process of bone formation and mostly incorporated into bone matrix with some escaping into the blood. Since the half-life in blood is relatively short (about 5 minutes) the osteocalcin level in blood reflects new protein synthesis and therefore its measurement provides a valuable tool for assessing skeletal metabolism. As a product unique to the osteoblast, it also represents the activity of the cell responsible for the formation of bone.

TEST PRINCIPLE

The Mouse Osteocalcin ELISA Kit is a two-site enzyme-linked immunosorbent assay (ELISA) for the measurement of osteocalcin in mouse serum, plasma or cell culture media. Two different goat polyclonal antibodies to mouse osteocalcin have been purified by affinity chromatography. The antibody which recognizes epitopes within the midregion/C-terminal portion of the peptide is biotinylated for capture. The other antibody which recognizes epitopes within the N- terminal region is conjugated with the enzyme horseradish peroxidase (HRP) for detection.

In a two-step reaction a sample containing mouse osteocalcin is first incubated with the biotinylated antibody in a streptavidin coated microtiter well. After washing the well to remove any unbound antibody and other components, the well is incubated with the HRP conjugated antibody. Osteocalcin contained in the sample is now immunologically bound by the capture antibody and the detection antibody to form a “sandwich” complex:

Well/Biotin Antibody — Mouse Osteocalcin — HRP Antibody

Following another wash, the enzyme antibody bound to the well is incubated with a substrate solution in a timed reaction and then measured in a spectrophotometric microtiter plate reader. The enzymatic activity of the antibody complex bound to the well is directly proportional to the amount of mouse osteocalcin in the sample. A standard curve is generated by plotting the absorbance versus the respective mouse osteocalcin concentration

ELISA Stop Solution**40-0030****11 mL**

One (1) bottle of 1 M sulfuric acid. This reagent may be stored at room temperature or at 2°C to 8°C and is stable until the expiration date on the kit.

Plate Sealer**10-2016**

Two (2) included in kit.

MATERIALS REQUIRED BUT NOT SUPPLIED IN KIT

- 1.0 mL and 2.0 mL volumetric pipettes for reconstituting standards and controls
- Precision pipets capable of delivering 20 µL, 25 µL, 50 µL, 100 µL and 200 µL
- Aluminum foil
- Automated microtiter plate washer OR
- Repeating dispenser for delivering 350 µL and suitable aspiration device
- Container for storage of wash solution
- Spectrophotometric microtiter plate reader capable of reading absorbance at 450 nm
- Deionized water
- Horizontal rotator capable of maintaining 180-220 RPM
- Timer

WARNINGS AND PRECAUTIONS

- Avoid contact with reagents containing TMB, hydrogen peroxide, or sulfuric acid (i.e. ELISA HRP Substrate and ELISA Stop Solution). TMB is dissolved in a solution which contains acetone, an irritant to skin and mucous membranes. In case of contact with any of these reagents, wash thoroughly with water. TMB is a suspected carcinogen. Use Good Laboratory Practices. Wash hands before eating. Do not eat, drink or smoke in the work area.
- Some of the reagents in this kit contain sodium azide. Sodium azide may react with lead or copper plumbing to form highly explosive metal azides. On disposal, flush with a large volume of water to prevent azide buildup (Manual Guide-Safety Management No. CDC-22, Center for Disease Control, Atlanta, Georgia, April 30, 1976).
- Testing should be performed in an area with adequate ventilation.
- Dispose of containers and unused contents in accordance with Federal, State and Local regulatory requirements.
- Wear suitable protective clothing, gloves, and eye/face protection when handling the contents of this kit.
- Wash hands thoroughly after handling.
- For additional information on hazard symbols, safety, handling and disposal of the components within this kit, please refer to the Safety Data Sheet (SDS) located at quidel.com.

Preparation and Storage

Prior to use allow all reagents to come to room temperature and mix by gentle swirling and inversion.

Reagents from different kit lot numbers should not be combined or interchanged.

Store the kit at 2°C to 8°C upon receipt. **Store the standards and controls at –20°C or below after reconstitution.** For the expiration date of the kit refer to the label on the kit box. All components are stable until this expiration date.

SPECIMEN COLLECTION

Measurement of the mouse osteocalcin concentration may be made on serum, plasma or cell culture media. Since serum and plasma samples are diluted 1:11 prior to assay only twenty microliters are required to assay the sample in duplicate. If obtaining serum, collect blood and allow it to clot at room temperature. Centrifuge

the sample and separate the serum, plasma, or media from the cells. Samples should be assayed immediately or stored frozen at -20°C or below. Avoid repeated freezing and thawing of specimens.

ASSAY PROCEDURE

1. **Dilute both controls and each serum or plasma sample 1:11 prior to assay.** (Standards do not require dilution and are ready-to-use after reconstitution.) For sample dilution pipette 20 μL of sample and 200 μL of zero standard into appropriately labeled tubes and vortex. Cell culture media samples may have to be diluted differently to obtain optimum results.
2. Place a sufficient number of Streptavidin Coated Strips in a holder to run osteocalcin standards, controls and unknown samples.
3. Pipet 25 μL of standard, diluted control, or diluted sample into the designated or mapped well. Freeze the remaining standards and controls except standard zero, as soon as possible after use.
4. Pipet 50 μL of Mouse Osteocalcin Biotinylated Antibody into each well and cover the plate with one plate sealer.
5. Incubate plate at room temperature for 1 hour on a horizontal rotator set at 180-220 RPM.
6. Remove the plate sealer. **Using an automated microtiter plate washer aspirate the contents of each well. Wash each well five times by dispensing 350 μL of working wash solution into each well and then completely aspirate the contents.** A suitable aspiration device may also be used.
7. Pipet 100 μL of Mouse Osteocalcin HRP conjugated antibody into each well.
8. Re-cover the plate with the plate sealer and aluminum foil to protect from exposure to light. Incubate at room temperature for 1 hour on a horizontal rotator set at 180-220 RPM.
9. Remove the aluminum foil and plate sealer. **Using an automated microtiter plate washer aspirate the contents of each well. Wash each well five times by dispensing 350 μL of working wash solution into each well and then completely aspirating the contents.** A suitable aspiration device may also be used.
10. Pipet 100 μL of ELISA HRP Substrate into each of the wells.
11. Re-cover the plate with the Plate Sealer and aluminum foil. Incubate at room temperature for 30 minutes on a horizontal rotator set at 180-220 RPM.
12. Remove the aluminum foil and plate sealer. Immediately pipet 50 μL of ELISA Stop Solution into each of the wells. Mix on a horizontal rotator for 1 minute.
13. Read the absorbance at 450 nm within 10 minutes in the microtiter plate reader against a reagent blank of 100 μL of Substrate and 50 μL of Stop Solution.

If dual wavelength correction is available set the instrument to Measurement wavelength to 450 nm and Reference wavelength to 595 nm to 650 nm.

PROCEDURAL NOTES

- It is recommended that all standards, controls and samples be assayed in duplicate. The average absorbance reading of each duplicate should then be used for data reduction and the calculation of results.
- Store light sensitive reagents (i.e. HRP Conjugated Antibody and ELISA HRP Substrate) in the original amber bottles or other suitable container which is well protected from light.
- Store any unused Streptavidin Coated Strips in the resealable aluminum pouch with desiccant to protect them from moisture.
- The sample and all reagents should be pipetted carefully to minimize air bubbles in the wells.
- The sequence and timing of each reagent addition is important as both the immunological and enzymatic reactions are in kinetic modes. The washing step is also an important part of the total assay procedure. **The use of an automated microtiter plate washer is strongly recommended.** All pipetting and washing steps should be performed such that the timing is as consistent as possible.
- Samples with values greater than the highest standard should be further diluted 1:10 with the 0 ng/mL Standard and reassayed. Multiply the result by 10. (See Limitations, # 1 and # 2)

- Plasma or cell culture media samples may contain fibrin clots or cellular debris. Freeze/thaw of plasma samples may accelerate clot formation. These samples must be centrifuged and decanted prior to assay to remove all particulate material which can cause random high non-specific binding on well surface.
- Rarely, upon opening the streptavidin plate, small white crystals may be observed in some of the wells. This is entirely cosmetic and will not affect the assay. This condition is reported by other kit manufacturers and results from the final stabilizing buffer used in the coating process.

CALCULATION OF RESULTS

The absorbance readings taken after the addition of the ELISA Stop Solution allow for the construction of a standard curve using the mouse osteocalcin standards contained in the kit. **Refer to the individual vial label for exact concentration.** The curve should be generated as follows:

1. Calculate the average absorbance for each pair of duplicate assay wells.
2. Subtract the average absorbance of the 0 ng/mL Standard from the average absorbance of all other readings to obtain corrected absorbance.
3. The standard curve is generated by plotting the corrected absorbance of each standard level on the ordinate against the standard concentration on the abscissa using linear-linear or log- log paper. Appropriate computer assisted data reduction programs may also be used for the calculation of mouse osteocalcin results.

The mouse osteocalcin concentration of the controls and samples are read directly from the standard curve using their respective corrected absorbance. If log-log graph paper or computer assisted data reduction programs utilizing logarithmic transformation are used, samples having corrected absorbance between the 0 ng/mL Standard and the next highest standard should be calculated by the formula:

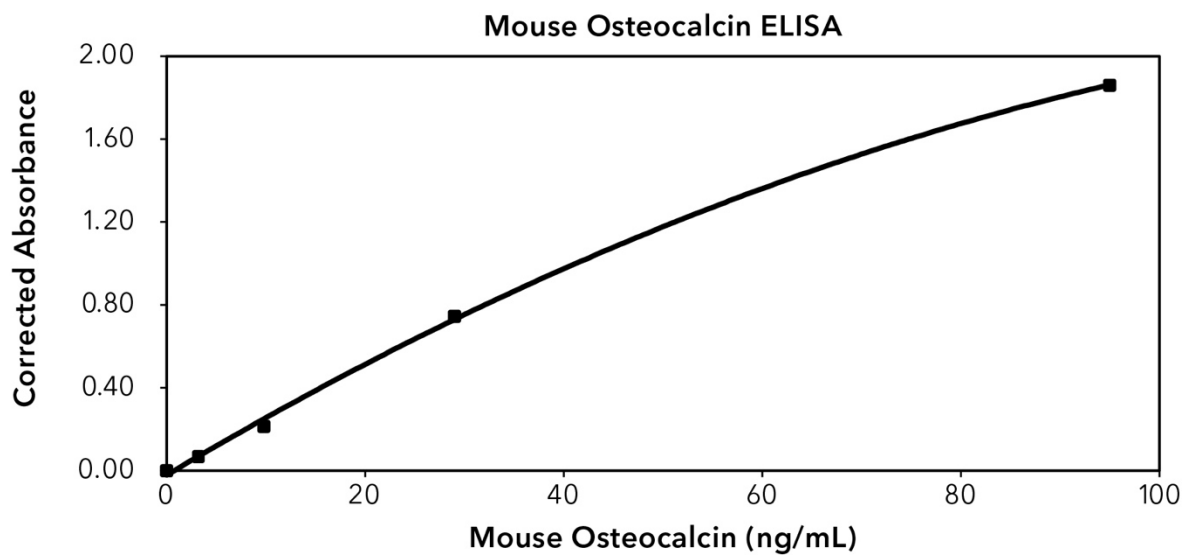
$$\text{Value of unknown} = \frac{\text{Corrected Absorbance (unknown)}}{\text{Corrected Absorbance (2}^{\text{nd}} \text{ Std.)}} \times \text{Value of the 2}^{\text{nd}} \text{ Std.}$$

To obtain the final mouse osteocalcin concentrations for controls and samples multiply the observed values by the dilution factor.

EXAMPLE DATA AND STANDARD CURVE

The following are representative examples of data and the resulting standard curve. **This curve should not be used in lieu of a standard curve run with each assay.**

ABSORBANCE—450 NM					
Well I.D.	ABS	Average ABS	Corrected ABS	Results ng/mL	Corrected Results ng/mL
Reagent Blank	0.000 0.000	0.000			
0 ng/mL	0.001 0.001	0.001	0.000		
3.2 ng/mL	0.069 0.069	0.069	0.068		
9.8 ng/mL	0.217 0.212	0.215	0.214		
29 ng/mL	0.746 0.747	0.746	0.745		
95 ng/mL	1.815 1.908	1.861	1.860		
Control I	0.562 0.563	0.562	0.561	22.5	247
Control II	1.474 1.470	1.472	1.471	72.3	795
Sample 1	0.370 0.378	0.374	0.373	15.7	173
Sample 2	0.153 0.146	0.149	0.148	6.9	76
Sample 3	0.991 1.015	1.002	1.001	44.5	489



LIMITATIONS OF THE PROCEDURE

- The lowest concentration of mouse osteocalcin measurable is 0.4 ng/mL (assay sensitivity) and the highest concentration of mouse osteocalcin measurable is the value of the highest standard.
- The reagents in this Mouse Osteocalcin ELISA kit have been optimized so that the high dose “hook effect” is not a problem for samples with elevated mouse osteocalcin values. Samples with mouse osteocalcin levels between the highest standard and 10,000 ng/mL will read greater than the highest standard and should be further diluted 1:10 with the 0 ng/mL Standard and reassayed for correct values.
- Grossly lipemic serum or plasma samples may affect the immunological response and it is recommended that results obtained with such samples be scrutinized accordingly.
- Differences in protein concentration and protein type between samples and standards in an immunoassay contribute to "protein effects" and dose biases. When measuring low protein concentration culture media samples against high protein concentration standards, it is recommended that like samples be assayed together in the same assay to minimize this bias.

QUALITY CONTROL

To assure the validity of the results each assay should include adequate controls with known levels of mouse osteocalcin. Quidel recommends that all assays include the laboratory's own mouse osteocalcin controls in addition to those provided with this kit.

PERFORMANCE CHARACTERISTICS

Sensitivity

The sensitivity of the mouse osteocalcin assay as determined by the 95% confidence limit on 20 duplicate determinations of the 0 ng/mL Standard is 0.4 ng/mL.

Precision

To assess intra-assay precision the mean and coefficient of variation were calculated from 20 duplicate determinations of two diluted samples each performed in a single assay.

Observed Mean Value (ng/mL)	Coefficient of Variation
25.3	3.7 %
76.1	2.3 %

To assess inter-assay precision the mean and coefficient of variation were calculated from duplicate determinations of two diluted samples performed in 20 assays.

Observed Mean Value (ng/mL)	Coefficient of Variation
25.4	6.1 %
81.5	4.1 %

Parallelism

Mouse serum and plasma samples were diluted with the 0 ng/mL Standard and assayed. Results in ng/mL are as follows:

SAMPLE	DILUTION	CORR. OBSERVED VALUE	CORR. EXPECTED VALUE	% O/E
1	1:11	186		
	1:22	108	93	116
	1:44	53	46	115
	1:88	25	23	109
2	1:11	778		
	1:22	400	389	103
	1:44	201	195	103
	1:88	110	97	113
3	1:11	888		
	1:22	437	444	98
	1:44	217	222	98
	1:88	106	111	95

Recovery

Two samples containing mouse osteocalcin were added to each other in varying ratios and assayed. Results, corrected for dilution, in ng/mL are as follows:

SAMPLE	RATIO	OBSERVED VALUE	EXPECTED VALUE	% O/E
A	100/0	42		
	75/25	217	240	90
	50/50	398	437	91
	25/75	592	635	93
	0/100	833		
B	100/0	37		
	75/25	220	237	93
	50/50	399	435	92
	25/75	589	634	93
	0/100	833		
C	100/0	49		
	75/25	225	245	92
	50/50	415	441	94
	25/75	618	637	97
	0/100	833		

Cross-reactivity

Human osteocalcin and rat osteocalcin were each diluted in an osteocalcin-free protein matrix and measured in the Mouse Osteocalcin ELISA Kit. The results show zero cross-reactivity from either of these species.

CORRELATION

A correlation study was performed with mouse serum and plasma samples comparing results obtained from the Mouse Osteocalcin ELISA Kit vs. results obtained from the Mouse Osteocalcin IRMA Kit (Immutopics, Inc.). A linear regression analysis gave the following results: $n = 91$, $ELISA = 1.02 \times IRMA - 0.5$; correlation coefficient: $(r) = 0.97$.

ASSISTANCE

To place an order or for technical support, please contact a Quidel Representative at 800.874.1517 (in the U.S.) or 858.552.1100 (outside the U.S.), Monday through Friday, from 8:00 a.m. to 5:00 p.m., Eastern Time. Orders may also be placed by fax at (740) 592-9820. For e-mail support contact customerservice@quidel.com or technicalsupport@quidel.com.

For services outside the U.S.A., please contact your local distributor. Additional information about Quidel, our products, and our distributors can be found on our website quidel.com.

REFERENCES

1. Desbois C, Hogue D, Karsenty G. "The mouse osteocalcin gene cluster contains three genes with two separate spatial and temporal patterns of expression." *J Biol Chem* 269:1183-1190, 1994.
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REF

60-1305 MicroVue Mouse Osteocalcin ELISA – 96 Test

RUO



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GLOSSARY

REF

Catalogue number

LOT

Batch code



Use by



Manufacturer



Temperature limitation



Consult e-labeling instructions for use

RUO

For Research use only



Contains sufficient for 96 determinations

CONT

Contents/Contains

CONTROL

Control
